

# Zebrafish Embryo Developmental Toxicity Assay

**Commonly used acronym:** ZEDTA

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## Organisation

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**Country** Belgium

**Geographical Area** Flemish Region

## SCOPE OF THE METHOD

<b>The Method relates to</b>	Human health
<b>The Method is situated in</b>	Regulatory use - Routine production, Translational - Applied Research
<b>Type of method</b>	In vitro - Ex vivo
<b>Species from which cells/tissues/organs are derived</b>	Zebrafish

<b>Type of cells/tissues/organs</b>	Embryo
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## DESCRIPTION

### Method keywords

development

toxicity

teratogen

screening

regulatory toxicology

malformations

drug development

chemicals

automated analysis

zebrafish

embryo

### Scientific area keywords

toxicity testing

drug screening

drug development

preclinical

teratogenicity

### Method description

In view of safety of pregnant women, a promising *in vitro* zebrafish embryo developmental toxicity assay has been developed to test pharmaceutical and chemical compounds for their teratogenic potential. The protocol deals with exposing zebrafish embryos to a range of compound concentrations at 28°C throughout organogenesis, i.e. from the gastrulation stage (5.25 hours post-fertilization [hpf]) up to 120 hpf. Morphological development is monitored at 5, 12, 24, 48, 72, 96 and 120 hpf. Larvae are evaluated for lethality in order to identify an LC25 (the compound concentration in which 25% lethality is observed) and morphological anomalies using a numerical scoring system to identify the NOAEL (no observed adverse effect level). These values are used to calculate the teratogenic index (LC25/NOAEL ratio) of each compound. If the teratogenic index is equal to or greater than 10 then the compound is classified as a teratogen, and if the ratio is less than 10 then the compound is classified as a non-teratogen. Currently the assay is optimized by including several skeletal endpoints after skeletal staining at 120 hpf and exogenous metabolic activation systems are developed to encompass the limited biotransformation capacity of the zebrafish embryos. Automation of the morphological scoring is also explored.

### **Lab equipment**

Stereomicroscope ;

Aquaria ;

Incubator.

### **Method status**

Still in development

Published in peer reviewed journal

## **PROS, CONS & FUTURE POTENTIAL**

### **Advantages**

Fast results ;

Medium-throughput ;

Cost effective ;

Limited compound requirements ;

Longitudinal follow-up.

## **Challenges**

Compound uptake (internal concentrations) ;

Limited biotransformation ;

Less morphological endpoints compared to the mammalian *in vivo* tests.

## **Modifications**

Skeletal staining methods and exogenous metabolic activation systems are currently developed to increase the sensitivity of the assay. The main focus is to reduce the number of false negative results.

## **Future & Other applications**

The main goal is to optimize and use the assay for (regulatory) developmental toxicity testing, but the assay could potentially also be used for chronic toxicity testing in the future.

## **REFERENCES, ASSOCIATED DOCUMENTS AND OTHER INFORMATION**

### **References**

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