

# Generation of brain organoid to study tumorigenesis

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## Organisation

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**Department** Biomedical Sciences

**Specific Research Group or Service**

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**Country** Belgium

**Geographical Area** Brussels Region

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## SCOPE OF THE METHOD

The Method relates to	Human health
The Method is situated in	Basic Research

<b>Type of method</b>	In vitro - Ex vivo
<b>Species from which cells/tissues/organs are derived</b>	human embryonic stem cells
<b>Type of cells/tissues/organs</b>	human pluripotent stem cells

## DESCRIPTION

### Method keywords

HUman brain organoids

tumor

human embryonic stem cell derived organoid model

### Scientific area keywords

Human Stem cells

tumorigenesis

3D organoid models

hESC-derived organoids

### Method description

This method uses human pluripotent stem cells (hPSCs), including embryonic and induced pluripotent stem cells, to generate cortical brain organoids and model brain tumorigenesis through targeted genetic manipulation. The aim is to create a physiologically relevant, *in vitro* 3D system that mimics early brain development and allows the study of cancer initiation and progression. Organoids are guided through neural induction and maturation using stage-specific culture conditions and matrix support, resulting in self-organized brain-like structures. Genetic alterations are introduced via lentiviral transduction of oncogenes and tumor suppressor knockouts to simulate tumor development. Fluorescent markers are used to trace cell origin and monitor growth dynamics over time. This model enables spatial and temporal investigation of tumor biology in a controlled setting and represents a promising tool for studying human brain cancer. Additionally, it has potential applications in modeling

other cancers, such as liver tumors, offering an ethical alternative to animal experimentation.

## **Lab equipment**

This method requires lab equipment including a biosafety cabinet for sterile cell culture work, an orbital shaker for organoid agitation, a fluorescent microscope for live imaging, and a flow cytometer to analyze and separate transduced cell populations. Access to lentiviral production facilities.

## **Method status**

History of use

Internally validated

Published in peer reviewed journal

## **PROS, CONS & FUTURE POTENTIAL**

### **Advantages**

- This method allows the simultaneous analysis of transformed and non-transformed tissues within the same organoid, enabling direct comparison in a controlled 3D environment.
- It supports high-throughput experimentation, as multiple organoids can be generated and analyzed in parallel.
- The approach offers a physiologically relevant model for tumor initiation and progression and reduces reliance on animal models.

### **Challenges**

- One major limitation is the variability between organoid batches, which can affect reproducibility and data interpretation.
- The method is also labor-intensive and requires significant hands-on time, technical expertise, and access to specialized equipment and reagents.

### **Modifications**

- Future optimization may focus on reducing batch-to-batch variability by standardizing organoid generation protocols or using automation.
- Advanced imaging techniques could enhance resolution and data richness.
- Improvements in viral transduction efficiency could be also improved.

## Future & Other applications

This method has broad potential beyond brain tumor modeling. It can be adapted to generate organoids from other tissues such as liver, pancreas, or colon, to study various types of cancer or organ-specific diseases. Additionally, it holds promise for drug screening, personalized medicine, and developmental biology studies.

## REFERENCES, ASSOCIATED DOCUMENTS AND OTHER INFORMATION

### References

doi:10.1038/s41592-018-0070-7; Doi: 10.1038/s41596-024-01107-7

### Associated documents

[Mutagensis induction in brain organoids.png](#)

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