Efficient CRISPR gene editing in primary cells and organoids using virus-like particles

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Organisation

Name of the organisation Katholieke Universiteit Leuven (KUL)
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Specific Research Group or Service
Laboratory of Respiratory Diseases and Thoracic Surgery (BREATHE)
Country Belgium
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Name of the organisation Katholieke Universiteit Leuven (KUL)
Department Pharmaceutical and Pharmacological Sciences
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Partners and collaborations

Katholieke Universiteit Leuven (KUL)

SCOPE OF THE METHOD

The Method relates to	Human health
The Method is situated in	Basic Research, Translational - Applied Research
Type of method	In vitro - Ex vivo
Specify the type of cells/tissues/organs	several primary cell models (rectal organoids, endothelial cells, epithelial cells, iPSCs, ...)

DESCRIPTION

Method keywords

organoids
genome engineering
gene editing
CRISPR-Cas9
CRISPR/Cas
virus-like particles
transduction
base edting

prime editing
knock out

Scientific area keywords

gene therapy
genetics
3D organoid models
human diseases
human health
human adult stem cells

Method description

Using the combination of advanced CRIPSR tools including several Cas orthologs, based editors (ABE, CBE, CGBE) and prime editing technologie with efficient delivery vehicles such as LV, AAV and virus-like particles (VLPs), our lab has become experienced with introducing or replacing precise edits in several primary cell models. For gene editing, VLPs are especially well suited because they link the efficacy of viral transduction with the delivery of RNP cargo, delivering a very transient dose of gene editing cargo. We have built VLPs harbouring several gene editing enzymes (Cre, several Cas variants, several base editors, several PE strategies), as well as reporter cargos (mNeonGreen, B-galactosidase, fLuc,...).

Lab equipment

BSL2 is required to work with cells from human origin and with particles capable of entering these cells.

Method status

Still in development
Internally validated

PROS, CONS & FUTURE POTENTIAL

Advantages

Efficient gene editing in hard-to-edit cell types using ultra transient exposure and therefore limiting risks on off-target editing. VLP preps can be ordered from the Leuven Viral Vector Core (https://gbiomed.kuleuven.be/english/corefacilities/LVVC/technology)

Challenges

VLP production is expensive and needs to be performed by experienced staff under highly standardised SOP's. We have large libraries of VLPs available and have shared many before under academic collaboration.

REFERENCES, ASSOCIATED DOCUMENTS AND OTHER INFORMATION

Links

Leuven Viral Vector Core

Other remarks

This work was performed in collaboration with the Leuven Viral Vector Core (Contact: lvvc@kuleuven.be).

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