

# In vitro generation of human hematopoietic cells

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#### Organisation

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# SCOPE OF THE METHOD

| The Method relates to                    | Human health                                 |
|--|--|
| The Method is situated in                | Basic Research                               |
| Type of method                           | In vitro - Ex vivo                           |
| Specify the type of cells/tissues/organs | Human hematopietic stem and progenitor cells |

# DESCRIPTION

#### Method keywords

human HPCs

in vitro differentiation of hematopoietic cells

OP9-coculture

MS5-coculture

ATO system

organoid culture FTOC

# Scientific area keywords

immune deficiency leukemia human hematopoiesis stem cells gene editing

## **Method description**

Better understanding of molecular mechanisms controlling both normal and malignant human hematopoiesis will lead to a more efficient therapy of immune deficiencies and lymphoid leukemias. Therefore, human hematopoietic progenitor cells (HPCs) are differentiated *in vitro* towards distinct hematopoietic lineages, with or without perturbation conditions such as gene targeting, viral transductions, specific compounds or blocking antibodies. Our lab has a broad expertise in the differentiation of human T cell progenitors, for which 3 different *in vitro* techniques are available:

 1) Fetal thymic organ cultures (FTOCs), using fetal thymic lobes from NOD/SCID mice as 3D-micro environment allowing human HPCs to differentiate towards T cells. ;
2) OP9-coculture system, using OP9 mouse stromal cells with or without specific Notch ligands as a 2D-layer to culture human HPCs on ;

3) Artificial Thymic Organoid (ATO) cultures, using Notch expressing MS5 mouse stromal cells in combination with human HPCs in 3D aggregates.

Furthermore, OP9- and MS5-cocultures are used in order to differentiate HPCs towards myeloid cells (dendritic cells, monocytes, granulocytes), B cells, NK cells and both erythrocytes and megakaryocytes. Differentiation of different cell types is determined using flow cytometry.

# Lab equipment

Biosafety cabinet level 2 ; Tissue culture incubator ; Centrifuge ; Flow cytometer.

#### Method status

Internally validated Published in peer reviewed journal

# **PROS, CONS & FUTURE POTENTIAL**

#### Advantages

These techniques allow to study normal or aberrant differentiation of human hematopoietic stem cells in conditions of genetic or other perturbations *in vitro*. It permits a kinetic and quantitative analysis of human hematopoietic differentiation which is difficult *in vivo*.

## Challenges

The challenge of *in vitro* differentiation systems is reproducing the *in vivo* environment in which different hematopoietic cells arise. Although FTOCs and the ATO system offer a close physiological background, the use of OP9 or MS5 stromal cells also allows us to generate distinct hematopoietic cells resembling their *in vivo* counterparts. Gene targeting in human HSCs is still inefficient.

#### Modifications

More efficient gene targeting in human HSCs is still desired for genetic studies, as well as further modifications that lead to a closer resemblance of the *in vivo* environment.

# **REFERENCES, ASSOCIATED DOCUMENTS AND OTHER INFORMATION**

#### References

Taghon T et al. Blood 2002; 99(4):1197-204. Schmitt TM et al. Immunity 2002; 17(6):749-56. Van de Walle I et al. Blood 2011; 117(17):4449-59. Seet CS et al. Nat Methods 2017; 14(5):521-530. Montel-Hagen A et al. Cell Stem Cell 2019; 24(3):376-389.

## Links

lab website

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